

Prevalence of *Taenia solium* cysticercosis in pigs entering the food chain in western Kenya

Lian Francesca Thomas^{1,2} · Leslie Jayne Stevenson Harrison³ · Philip Toye² · William Anson de Glanville^{1,2} · Elizabeth Anne Jesse Cook^{1,2} · Claire Njeri Wamae^{4,5} · Eric Maurice Fèvre^{2,6}

Received: 29 September 2015 / Accepted: 27 October 2015 / Published online: 18 November 2015
© The Author(s) 2015. This article is published with open access at Springerlink.com

Abstract Three hundred forty-three pigs slaughtered and marketed in western Kenya were subjected to lingual examination and HP10 Ag-ELISA for the serological detection of *Taenia solium* antigen. When estimates were adjusted for the sensitivity and specificity of the diagnostic assays, prevalence of *T. solium* cysticercosis estimated by lingual exam and HP10 Ag-ELISA was between 34.4 % (95 % confidence interval (CI) 19.4–49.4 %) and 37.6 % (95 % CI 29.3–45.9 %), respectively. All pigs, however, were reported to have passed routine meat inspection. Since *T. solium* poses a serious threat to public health, these results, if confirmed, indicate that the introduction of control strategies may be appropriate to ensure the safety of pork production in this region.

Keywords *Taenia solium* · Cysticercosis · Zoonotic · Epidemiology · Public health · Kenya

✉ Eric Maurice Fèvre
Eric.Fevre@liverpool.ac.uk

¹ Centre for Immunity, Infection and Evolution, Institute for Immunology and Infection Research, School of Biological Sciences, Ashworth Labs, University of Edinburgh, West Mains Rd, Edinburgh EH9 3JT, UK

² International Livestock Research Institute, PO Box 30709, Nairobi 00100, Kenya

³ Royal (Dick) School of Veterinary Medicine, University of Edinburgh, Easter Bush, Edinburgh, Scotland, UK

⁴ Centre for Microbiology Research, Kenya Medical Research Institute, Nairobi, Kenya

⁵ School of Health Sciences, Mount Kenya University, Thika, Kenya

⁶ Institute of Infection and Global Health, University of Liverpool, Leahurst Campus, Neston CH64 7TE, UK

Introduction

Cysticercosis, infection with the intermediate stage of the tapeworm *Taenia solium*, is a zoonotic disease of public health and economic importance and is widespread in Africa, Asia and Latin America (Coral-Almeida et al. 2015). It is a leading cause of acquired epilepsy in humans and a priority for control under the 2012 road map for control of neglected tropical diseases ‘NTD roadmap’ (World Health Organization 2012).

The epidemiological picture for this parasite in sub-Saharan Africa (SSA) is becoming clearer, and it appears that, apart from predominately non-pork-eating areas, *T. solium* is present in practically all countries in the region (Assana et al. 2013). If validated control programmes are to be rolled out as required by the NTD roadmap (World Health Organization 2012), epidemiological data are required to identify areas both in need of control programmes and as baseline data prior to the roll-out of programmes.

Pig keeping in Kenya generally occurs as a small-holder industry with between 2 and 10 pigs per farm with many pigs kept under a free-range system (Kagira et al. 2010), under which they roam over large distances scavenging for food (Thomas et al. 2013). The pigs raised in this way are sold into the local market, either directly to butchers or via traders who travel from farm to farm purchasing pigs (Kagira et al. 2009). Official inspection of these pigs is required by law at the slaughter facility under the Meat Control Act (Government of Kenya 1977). The ad hoc arrangement of the market operating in a largely informal value chain, in conjunction with an understaffed meat inspectorate, however, allows a proportion of pork to enter the food chain without inspection (Kagira et al. 2009). This puts the population at risk from various food-borne diseases, including the parasite *T. solium*.

This study investigated the prevalence of *T. solium* cysticercosis in pigs entering the food chain in western Kenya as a first step to understanding the burden of this parasite in this region.

Method and materials

The study site, shown in Fig. 1, comprised the 10 divisions with the highest pig populations within a 45-km circumference of Busia town in what was previously western Kenya (now comprising Busia and parts of Siaya Counties). The study area is described in greater detail by Thomas et al. (2013).

A total of 343 sera samples from pigs were collected from porcine slaughter facilities, registered with the district veterinary office, between February and August 2010. A minimum sample size of 319 pigs was calculated to be required, using WinEpiscope 2.0 (Thrusfield et al. 2001) and based upon a maximum assumed *T. solium* prevalence of 14 % (Githigia et al. 2007) with 5 % precision and 99 % confidence level. Slabs were visited on the day of the highest throughput as

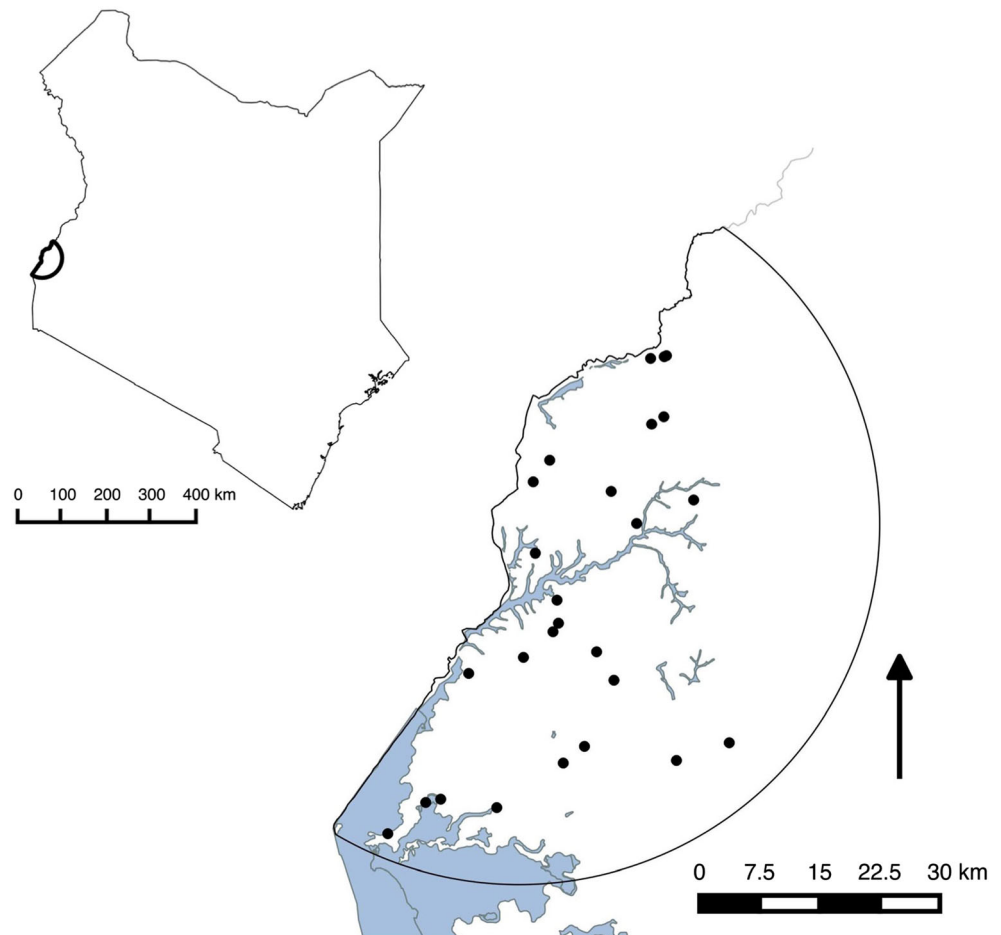
identified by the meat inspector or slab owner, and all pigs being slaughtered on the day of visit were sampled. Facilities were revisited until a quota proportional to the percentage of the total pig population found in that division had been filled.

Pigs were restrained with a pig snare behind the canine teeth, and a lingual palpation was carried out using a short stick to open the mouth and a cotton swab to protract the tongue. The ventral surface of the tongue was inspected for the presence of cysticerci (Mutua et al. 2007).

Anterior vena cava blood samples (Muirhead 1981) were collected in BD Vacutainer® 10-ml plain tubes and transported to the laboratory on ice, where they were centrifuged at 3000 rpm for 20 min at room temperature. Sera were then separated into two aliquots in 2-ml labelled cryovials and stored at -40°C until they were transported on dry ice to the International Livestock Research Institute (ILRI) facility in Nairobi where they were stored at -80°C for between 2 and 7 months prior to laboratory analysis.

Meat inspection results were obtained by contacting the meat inspector on duty for that slaughter facility at the end of the day and requesting his or her report on all pigs slaughtered. The investigators for this study did not interfere

Fig. 1 Map of Kenya indicating the location of the study site. The enlargement of the study site depicts the location of slaughter facilities (black dots), and the black arrow indicates north



in any way with the process of meat inspection as we wished to obtain a true reflection of the activities carried out. A further communication was made with all the meat inspectors involved at the conclusion of the study during which qualitative data were gathered on carcass condemnation for any reason during the time that this survey was taking place. Inspection of pigs at slaughter should be conducted as per the Kenya Meat Control Act (Government of Kenya 1977) which calls for palpation and incision of the tongue, visual inspection and incision of the heart and visual inspection of all exposed muscles, especially the neck, loin, ham and fleshy part of the diaphragm. A carcass found to be infected with any number of cysticerci is judged to be unfit for human consumption, and the whole carcass is condemned. No cysts were collected in this study for morphologic or PCR confirmation.

Laboratory analysis was carried out at ILRI, Nairobi. The HP10 Ag-ELISA (Harrison et al. 1989), modified as described by Harrison et al. (2005) and Krecek et al. (2008, 2011) was used for the detection of active cysticercosis.

Five known negative and two known positive reference controls were run on every plate in the screen.

Plates were corrected for plate-to-plate and day-to-day variations, using a correction factor determined from the first plate of the screening run as described by Harrison et al. (2005).

Cut-off values were determined using the mean corrected optical density (OD) of all negative controls run during the full screen plus three standard deviations ($OD=0.176$). Any serum sample with a corrected OD value over this cut-off was counted as being positive.

Results were entered into Microsoft Access 2007 database and were analysed using the 'R' environment for statistical computing (R Development Core Team 2005). In order to take into account the sensitivity (Se) and specificity (Sp) of the diagnostic assay, the 'epi.prev' function in epiR (Stevenson et al. 2013) was used to adjust the prevalence estimate. The Se/Sp of the HP10 Antigen ELISA has been estimated to be Se 89.5 % (95 % confidence interval (CI) 82.3–94.2) and Sp 74 % (95 % CI 56.6–87.6) (Porphyre et al. 2015), and lingual palpation was estimated to be Se 16.1 % (95 % CI 5–34 %) and Sp 100 % (97.5 % one-sided CI 90–100 %) (Dorny et al. 2004). No adjustment was made for clustering within the population as although sampling took place at a discrete number of slaughter facilities, all pigs originated from separate farms.

Results

Three hundred forty-three (343) pigs were sampled. One hundred seventy-one (171) pigs were found to be positive by HP10 Ag-ELISA with only nineteen (19) detected to be positive by lingual palpation (see Table 1). The overall prevalence for the study region, adjusting for the current estimates of diagnostic

test Se/Sp, is estimated to be 37.6 % (95 % CI 29.3–45.9 %) by HP10 Ag-ELISA and 34.4 % (95 % CI 19.4–49.4 %) by lingual palpation. Reports obtained from the meat inspectors indicated that no pigs with cysticercosis infections were detected at meat inspection and, in fact, no pig carcass or part thereof had been condemned for any reason (including but not limited to cysticercosis) during the time of the study. It is worth noting that on many occasions, no inspector was observed at the slaughter facility. At these times, pig carcasses were seen to leave the premises without inspection. When inspectors were contacted, we were informed that meat had been inspected at the butchery but we are unable to confirm this.

Discussion

This study has found a high prevalence of pigs testing positive to the HP10 Ag-ELISA, suggestive of viable *T. solium* cysticercosis infections, which places western Kenya within the prevalence range (4–567 %) detected in other endemic areas within Africa (Assana et al. 2013). Pork consumption in the developing countries has been increasing at an astonishing rate (Delgado 2003), and, in keeping with this trend, pig keeping has become a popular small-holder activity for low-income families in western Kenya (Kagira et al. 2010). Zoonotic infections, such as *T. solium*, however, may be a key limiting factor in the economic viability of small-holder livestock keeping (Grace et al. 2012).

The large difference seen between the 'crude' prevalence estimates obtained with lingual palpation and HP10 Ag-ELISA (see Table 1) is a direct result of the relative Se/Sp of the two tests. Lingual palpation is a quick, low-cost diagnostic test which has been reported to be used routinely by farmers in Peru to screen their own animals (Gonzalez et al. 1990). It is highly specific for *T. solium* but has very low sensitivity (Dorny et al. 2004; Krecek et al. 2008, 2011), and although it may have a role as a rapid screening tool for endemic areas, it has limited applicability in prevalence studies. The HP10 Ag-ELISA has a much higher sensitivity, though poorer specificity, than lingual palpation. False positives may be detected, including potential cross reactivity with *Taenia hydatigena*, although the prevalence of this parasite is assumed to be low in the pig population of sub-Saharan Africa (Dorny et al. 2004). Adjusting the crude prevalence figures to take these diagnostic characteristics into account provides us with a more 'true' impression of the likely parasite prevalence in this region.

The 95 % confidence intervals surrounding these adjusted prevalence estimates are wide and indicate the uncertainty still surrounding the performance of both diagnostic assays for *T. solium* (Porphyre et al. 2015). Here, the estimates of Se/Sp used were sourced from studies utilising non-gold-standard techniques (Dorny et al. 2004; Krecek et al. 2008, 2011; Porphyre et al. 2015). Further work is planned to further

Table 1 Prevalence of *T. solium* Cysticercosis in pigs in Western Kenya as determined by lingual palpation and HP10 Ag-ELISA

Division slaughter	Total pigs	Lingual positive	% Prevalence lingual palpation ^a (95 % CI)	Adjusted prevalence lingual palpation	HP10 positive	% Prevalence HP10 Ag-ELISA ^a (95 % CI)	Adjusted prevalence HP10 Ag-ELISA
Amagoro	10	0	0 (0–41.2)		4	40.0 (12.2–73.8)	
Amakura	27	0	0 (0–18.3)		12	44.4 (25.5–64.7)	
Budalangi	19	3	15.8 (3.4–39.6)		15	78.9 (31.9–71.33)	
Butula	24	2	8.3 (1.0–27.0)		12	50.0 (29.1–70.9)	
Chakol	14	0	0 (0–31.9)		8	57.1 (28.9–82.3)	
Funyula	69	4	5.8 (1.6–14.2)		43	62.3 (49.8–73.7)	
Matayos	42	2	4.8 (0.6–16.2)		23	54.8 (38.7–70.2)	
Nambale	105	5	4.8 (1.6–10.8)		43	41.0 (31.5–51.0)	
Ugunja	7	1	14.3 (0.4–57.9)		2	28.6 (3.7–71)	
Ukwala	26	2	7.7 (0.9–25.1)		9	34.6 (17.2–55.7)	
Total	343	19	5.5 (3.4–8.5)	34.4 (19.4–49.4)	171	49.9 (44.4–55.3)	37.6 (29.3–45.9)

^a Crude prevalence not adjusted for Se/Sp

explore diagnostic test performance using large-scale fine dissection of carcasses and cyst enumeration as a ‘gold-standard’ diagnostic.

That the pigs sampled in this study were entering the food chain, with no reported condemnation of carcasses by meat inspectors, raises important concerns for public health in this region. Consumption of this infected meat can lead to taeniasis, with any one *T. solium* carrier intermittently shedding thousands of eggs with the potential to cause neurocysticercosis in those people who may unwittingly ingest them (Garcia et al. 2003). A cornerstone of cysticercosis control is preventing infected pork meat from entering the food chain, and our data will therefore be valuable in designing interventions to improve food safety in this setting.

Various options are available for the control of this important zoonotic parasite although evidence for their sustained success is thus far lacking (Carabin and Traoré 2014). Improvement of the meat inspection services has been partially responsible for control of the parasite in Europe (Gemmill 1987). Meat inspection has, however, a notoriously poor sensitivity for the detection of cysticerci (Dorny et al. 2004; Boa et al. 2002; Gonzalez et al. 1990), as evidenced by the data presented here. Resource constraints leading to an understaffed meat inspectorate with insufficient powers to enforce legislation, particularly the condemnation of pigs, also seriously limits the success of meat inspection as a form of control (Thomas 2015).

The resource constraints experienced by the meat inspectorate in western Kenya were illustrated by the many occasions on which no inspector was observed at the slaughter facility. A similar situation was observed in another survey within our study area which found that a meat inspector visited only 84 % (95 % CI 81–87 %) of pig slaughter facilities (Cook 2014).

Should a better resourced inspectorate, and strict enforcement of legislation, be achieved, farmers may be encouraged to utilise

the informal slaughter and marketing routes that already operate within western Kenya. This is evidenced by the 3/63 pig farmers who reported utilising informal slaughter in a cross-sectional survey carried out within the same study site (Thomas 2014). It is important therefore that whichever strategy is adopted, all actors in the pork value chain, be they farmers, government institutions or consumers, are sufficiently incentivised to participate in control so that infected pigs are not moved from the formal market into clandestine informal value chains.

Conclusion

This study indicates that *T. solium* may be endemic in the porcine population of western Kenya but does not appear to be detected at meat inspection. This raises a very real public health concern for the communities who consume this pork and places the economic security of the industry at jeopardy. This area of Kenya should be considered for the roll-out of validated control programmes in accordance with the 2012 NTD roadmap.

Acknowledgments We would like to acknowledge Tim Strugnell from the Veterinary Laboratories Agency (UK) for supplying the negative control sera and all members of the PAZ field and laboratory teams, in particular James Akoko, Omoto Lazarus, Velma Kivali and Alice Kiyonga. The BBSRC supported LFT and WAdG, and the MRC supported EAJC. The People, Animals and their Zoonoses (PAZ) project and the Busia laboratory was supported by the Wellcome Trust (085308) and through a Strategic Award to the Centre for Immunity, Infection and Evolution (095831). This work was also supported by the Biotechnology and Biological Sciences Research Council, the Department for International Development, the Economic and Social Research Council, the Medical Research Council, the Natural Environment Research Council and the Defence Science and Technology Laboratory, under the Zoonoses and Emerging Livestock Systems (ZELS) programme, grant reference BB/L019019/1. The University of Edinburgh Innovation Initiative Grant fund is acknowledged for financially supporting the sample collection and the

Association for Strengthening Agricultural Research in East and Central Africa (ASARECA) for supporting the ELISA testing. We would also like to thank those pig farmers, traders, butchers and meat inspectors who so willingly participated in this study.

Compliance with ethical standards

Statement on animal rights Ethical approval for the sample collection from animals was granted by the Animal Welfare and Ethical Review Body (AWERB) at The Roslin Institute, University of Edinburgh (approval number AWA004 Broonsvort). Sampling of privately owned domestic pigs presented to slaughter houses was carried out by trained veterinarians or animal health assistants after obtaining verbal informed consent from the owners of those pigs. Blood sampling was undertaken according to the guidelines provided by the National Centre for the Replacement, Refinement and Reduction of Animals in Research (<http://www.nc3rs.org.uk/bloodsamplingmicrosite/page.asp?id=346>).

Conflict of interest Leslie J.S. Harrison is the Editor-in-chief of Tropical Animal Health and Production. No other conflict of interest exists.

Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

References

- Assana, E., Lightowers, M.W., Zoli, A.P., Geerts, S., 2013. *Taenia solium* taeniosis/cysticercosis in Africa: risk factors, epidemiology and prospects for control using vaccination. *Veterinary Parasitology* 195, 14–23.
- Boa, M.E., Kassuku, A.A., Willingham, A.L., Keyyu, J.D., Phiri, I.K., Nansen, P., 2002. Distribution and density of cysticerci of *Taenia solium* by muscle groups and organs in naturally infected local finished pigs in Tanzania. *Veterinary Parasitology* 106, 155–164.
- Carabin, H., Traoré, A.A., 2014. *Taenia solium* taeniosis and cysticercosis control and elimination through community-based interventions. *Current tropical medicine reports* 1, 181–193.
- Cook, E.A.J., 2014. The epidemiology of zoonoses in slaughterhouse workers in western Kenya (PhD Thesis, University of Edinburgh).
- Coral-Almeida, M., Gabriël, S., Abatih, E.N., Praet, N., Benitez, W., Dorny, P., 2015. *Taenia solium* Human Cysticercosis: A Systematic Review of Sero-epidemiological Data from Endemic Zones around the World. *PLoS Negl Trop Dis* 9, e0003919.
- Delgado, C.L., 2003. Rising consumption of meat and milk in developing countries has created a new food revolution. *The Journal of Nutrition* 133, 3907S–3910S.
- Dorny, P., Phiri, I.K., Vercauteren, J., Gabriel, S., Willingham, A.L., Brandt, J., Victor, B., Speybroeck, N., Berkvens, D., 2004. A Bayesian approach for estimating values for prevalence and diagnostic test characteristics of porcine cysticercosis. *International Journal for Parasitology* 34, 569–576.
- García, H.H., Gonzalez, A.E., Evans, C.A.W., Gilman, R.H., 2003. *Taenia solium* cysticercosis. *The Lancet* 362, 547–556.
- Gemmell, M., 1987. A critical approach to the concepts of control and eradication of echinococcosis/hydatidosis and taeniosis/cysticercosis. *International Journal for Parasitology* 17, 465–472.
- Githigia, S.M., Murekefu, A., Otieno, R.O., 2007. Prevalence of porcine cysticercosis and risk factors for *Taenia solium* taeniosis in Funyula Division of Busia District, Kenya. *Kenya Veterinarian* 29, 37–39.
- Gonzalez, A.E., Cama, V., Gilman, R.H., Tsang, V.C.W., Pilcher, J.B., Chavera, A., Castro, M., Montenegro, T., Verastegui, M., Miranda, E., Bazalar, H., 1990. Prevalence and Comparison of Serologic Assays, Necropsy, and Tongue Examination for the Diagnosis of Porcine Cysticercosis in Peru. *Am J Trop Med Hyg* 43, 194–199.
- Government of Kenya, 1977. Meat Control Act.
- Grace, D., Mutua, F., Ochungo, P., Kruska, R., Jones, K., Brierley, L., Lapar, L., Said, M., Herrero, M., Phuc, P., 2012. Mapping of poverty and likely zoonoses hotspots.
- Harrison, L.J.S., Garate, T., Bryce, D.M., Gonzalez, L.M., Foster-Cuevas, M., Wamae, L.W., Onyango-Abuje, J.A., Parkhouse, R.M.E., 2005. Ag-ELISA and PCR for monitoring the vaccination of cattle against *Taenia saginata* cysticercosis using an oncospherical adhesion protein (HP6) with surface and secreted localization. *Tropical animal health and production* 37, 103–120.
- Harrison, L.J.S., Joshua, G.W.P., Wright, S.H., Parkhouse, R.M.E., 1989. Specific detection of circulating surface/secreted glycoproteins of viable cysticerci in *Taenia saginata* cysticercosis. *Parasite Immunology* 11, 351–370.
- Kagira, J.M., Kanyari, P.N., Maingi, N., Githigia, S.M., Ng'ang'a, J.C., Karuga, J.W., 2010. Characteristics of the smallholder free-range pig production system in western Kenya. *Tropical animal health and production* 42, 865–873.
- Kagira, J.M., Maingi, N., Kanyari, P.W.N., Githigia, S.M., Ng'ang'a, J.C., Gachohi, J.M., 2009. Characteristics of pig trade in low income settings in Busia District, Kenya. *The Tropical Veterinarian* 26, 1–9.
- Krecek, R.C., Michael, L.M., Schantz, P.M., Ntanjana, L., Smith, M.F., Dorny, P., Harrison, L.J.S., Grimm, F., Praet, N., Willingham, A.L., 2008. Prevalence of *Taenia solium* cysticercosis in swine from a community-based study in 21 villages of the Eastern Cape Province, South Africa. *Veterinary Parasitology* 154, 38–47.
- Krecek, R.C., Michael, L.M., Schantz, P.M., Ntanjana, L., Smith, M.F., Dorny, P., Harrison, L.J.S., Grimm, F., Praet, N., Willingham, A.L., 2011. Corrigendum to “Prevalence of *Taenia solium* cysticercosis in swine from a community-based study in 21 villages of the Eastern Cape Province, South Africa” [*Vet. Parasitol.* 154 (2008) 38–47]. *Veterinary Parasitology*.
- Muirhead, M.R., 1981. Blood sampling in pigs. *In Practice* 3, 16–20.
- Mutua, F.K., Randolph, T.F., Arimi, S.M., Kitale, P.M., Githigia, S.M., Willingham, A.L., Njeruh, F.M., 2007. Palpable lingual cysts, a possible indicator of porcine cysticercosis, in Teso District, Western Kenya. *Journal of Swine Health and Production* 15, 206.
- Porphyre, V., Betson, M., Rabezanaahary, H., Mboussou, Y., Zafindraibe, N., Rasamoelina-Andriamanivo, H., Costard, S., Pfeiffer, D., Michault, A., 2015. *Taenia solium* porcine cysticercosis in Madagascar: Comparison of immuno-diagnostic techniques and estimation of the prevalence in pork carcasses traded in Antananarivo city. *Veterinary Parasitology* In Press.
- R Development Core Team, 2005. R: A language and environment for statistical computing. R Foundation for Statistical Computing Vienna.
- Stevenson, M., Nunes, T., Sanchez, J., Thornton, R., Reiczigel, J., Robison-Cox, J., Sebastiani, P., 2013. epiR: An R package for the analysis of epidemiological data. R package version 0.9-43.
- Thomas, L., de Glanville, W., Cook, E., Fevre, E., 2013. The spatial ecology of free-ranging domestic pigs (*Sus scrofa*) in western Kenya. *BMC veterinary research* 9, 46.
- Thomas, L.F., 2014. Epidemiology of *Taenia solium* cysticercosis in western Kenya (PhD Thesis, University of Edinburgh).
- Thomas, L.F., 2015. Landscape Analysis: Control of *Taenia solium*. World Health Organization, Geneva, Switzerland.

Thrusfield, M., Ortega, C., De Blas, I., Noordhuizen, J.P., Frankena, K., 2001. WIN EPISCOPE 2.0: improved epidemiological software for veterinary medicine. *Veterinary Record* 148, 567.

World Health Organization, 2012. *Accelerating Work to Overcome the Global Impact of Neglected Tropical Diseases: a Roadmap for Implementation*. WHO, Geneva.